

- [54] **MONOCLONAL ANTIBODY TO HUMAN HELPER T CELLS AND METHODS OF PREPARING SAME**
- [75] **Inventors:** Patrick C. Kung, Bridgewater; Gideon Goldstein, Short Hills, both of N.J.
- [73] **Assignee:** Ortho Pharmaceutical Corporation, Raritan, N.J.
- [21] **Appl. No.:** 33,639
- [22] **Filed:** Apr. 26, 1979
- [51] **Int. Cl.³** A61K 39/395; C12N 5/00; C12N 5/02; C12N 15/00; C12Q 1/00; G01N 33/48; G01N 33/68; G01N 33/96
- [52] **U.S. Cl.** 424/85; 260/112 R; 424/101; 424/177; 435/7; 435/172; 435/240; 435/241; 436/548
- [58] **Field of Search** 424/8, 12, 85, 88, 89, 424/101, 1, 177; 435/1, 7, 172, 240, 241; 260/112 R, 112 B; 436/548

[56] **References Cited**

U.S. PATENT DOCUMENTS

4,196,265 4/1980 Koprowski 435/2

OTHER PUBLICATIONS

Kennett, SSIE Data Bank Abstract of Grant, 5/77-4/79, Ab. No. ICA 189303.
 Herzenberg et al., SSIE Data Bank, Abstract of Grant, 9/77-7/79, Ab. No. ICA468120.
 Melchers (Ed) Lymph. Hybridoms, 2nd. Workshop on

Functional Properties of Tumors of T & B Lymph., Springer Verlag, Berlin, N.Y., 1978, pp. IX-XVIII.
 Yelton et al., Supra, pp. 1-7.
 Trucco et al., Supra, pp. 66-69.
 Levy et al., Supra, pp. 164-169.
 Hammerling et al., Supra, pp. 100-106.
 Iverson et al., Supra, pp. 192-194.
 Simpson et al., Supra, pp. 195-202.
 Ruddle et al., Supra, pp. 203-211.
 Taniguchi et al., Supra, pp. 212-216.
 Osborne et al., Supra, pp. 217-220.
 Saravia et al., Supra, pp. 224-231.
 Fox, C & E News, Jan. 1, 1979, pp. 15-17.
 White et al., J. Exp. Med., vol. 148, Sep. 1, 1978, pp. 664-673.
 Herzenberg et al., from Wier (Ed.) Handbook of Exptl. Immunol., Blackwell, London, 3rd Ed., 1978, pp. 25.1-25.7.

Primary Examiner—Anna P. Fagelson

Attorney, Agent, or Firm—Geoffrey G. Dellenbaugh

[57] **ABSTRACT**

Hybrid cell line for production of monoclonal antibody to an antigen found on all normal human helper T cells. The hybrid is formed by fusing splenocytes from immunized CAF₁ mice with P3X63Ag8U1 myeloma cells. Diagnostic and therapeutic uses of the monoclonal antibody are also disclosed.

11 Claims, 7 Drawing Figures

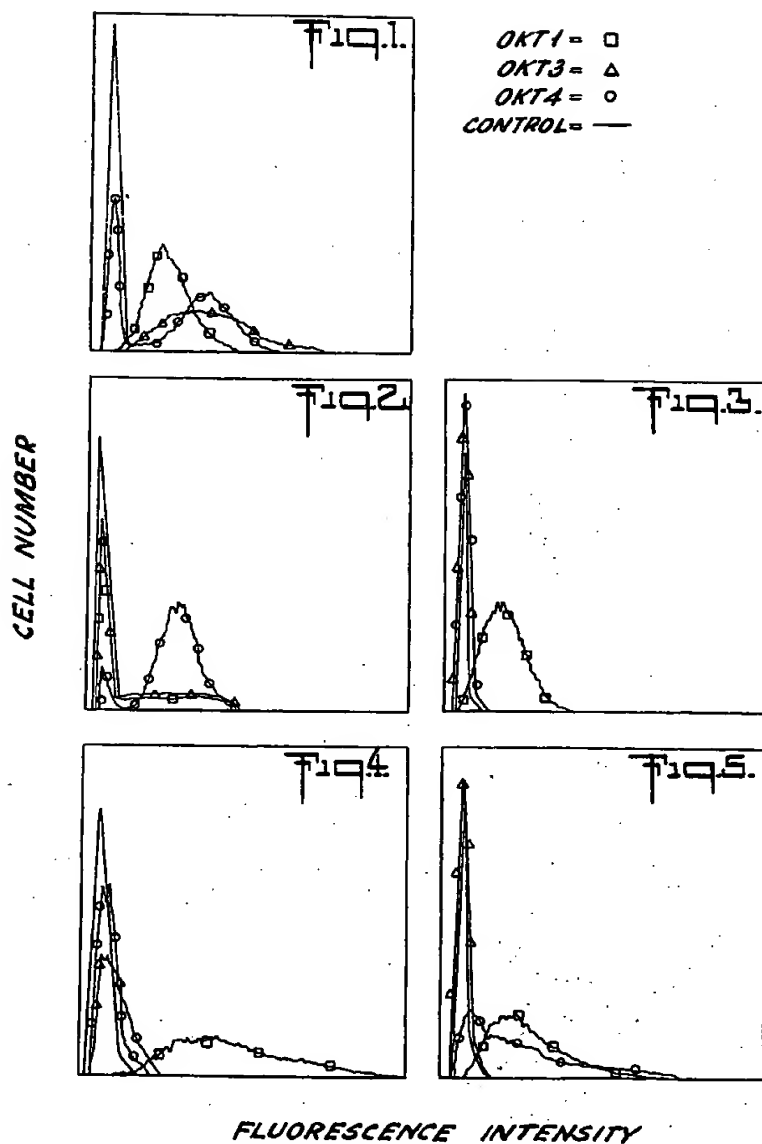


Fig. 6.

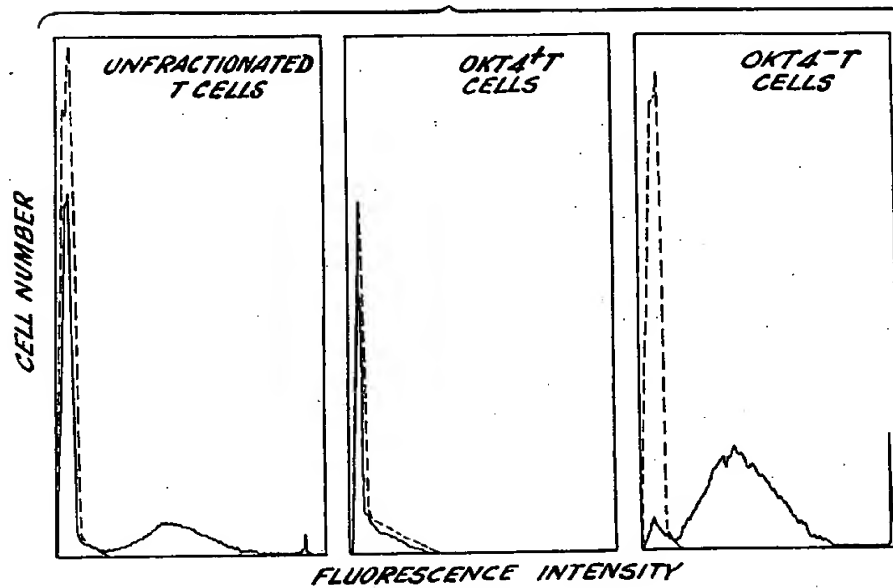
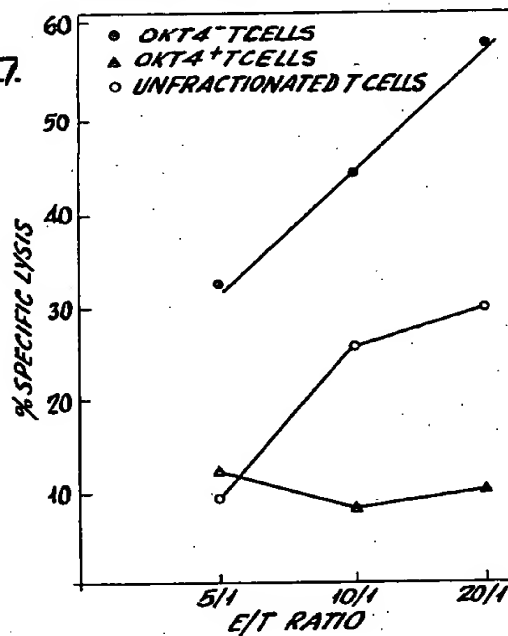


Fig. 7.



MONOCLONAL ANTIBODY TO HUMAN HELPER T CELLS AND METHODS OF PREPARING SAME

FIELD OF THE INVENTION

This invention relates generally to new hybrid cell lines and more specifically to hybrid cell lines for production of monoclonal antibody to an antigen found on all normal human helper T cells, to the antibody so produced, and to therapeutic and diagnostic methods and compositions employing this antibody.

DESCRIPTION OF THE PRIOR ART

The fusion of mouse myeloma cells to spleen cells from immunized mice by Kohler and Milstein in 1975 [Nature 256, 495-497 (1975)] demonstrated for the first time that it was possible to obtain a continuous cell line making homogeneous (so-called "monoclonal") antibody. Since this seminal work, much effort has been directed to the production of various hybrid cells (called "hybridomas") and to the use of the antibody made by these hybridomas for various scientific investigations. See, for example, *Current Topics in Microbiology and Immunology*, Volume 81-"Lymphocyte Hybridomas", F. Melchers, M. Potter and N. Warner, Editors, Springer-Verlag, 1978, and references contained therein; C. J. Barnstable, et al., *Cell*, 14, 9-20 (May, 1978); P. Parham and W. F. Bodmer, *Nature* 276, 397-399 (November, 1978); *Handbook of Experimental Immunology*, Third Edition, Volume 2, D. M. Wier, Editor, Blackwell, 1978, Chapter 25; and *Chemical and Engineering News*, Jan. 1, 1979, 15-17. These references simultaneously indicate the rewards and complications of attempting to produce monoclonal antibody from hybridomas. While the general technique is well understood conceptually, there are many difficulties met and variations required for each specific case. In fact, there is no assurance, prior to attempting to prepare a given hybridoma, that the desired hybridoma will be obtained, that it will produce antibody if obtained, or that the antibody so produced will have the desired specificity. The degree of success is influenced principally by the type of antigen employed and the selection technique used for isolating the desired hybridoma.

The attempted production of monoclonal antibody to human lymphocyte cell-surface antigens has been reported only in a few instances. See, for example, *Current Topics in Microbiology and Immunology*, *ibid*, 66-69 and 164-169. The antigens used in these reported experiments were cultured human lymphoblastoid leukemia and human chronic lymphocytic leukemia cell lines. Many hybridomas obtained appeared to produce antibody to various antigens on all human cells. None of the hybridomas produced antibody against a predefined class of human lymphocytes.

It should be understood that there are two principal classes of lymphocytes involved in the immune system of humans and animals. The first of these (the thymus-derived cell or T cell) is differentiated in the thymus from haemopoietic stem cells. While within the thymus, the differentiating cells are termed "thymocytes." The mature T cells emerge from the thymus and circulate between the tissues, lymphatics, and the bloodstream. These T cells form a large proportion of the pool of recirculating small lymphocytes. They have immunological specificity and are directly involved in cell-mediated immune responses (such as graft rejection) as effector cells. Although T cells do not secrete humoral

antibodies, they are sometimes required for the secretion of these antibodies by the second class of lymphocytes discussed below. Some types of T cells play a regulating function in other aspects of the immune system. The mechanism of this process of cell cooperation is not yet completely understood.

The second class of lymphocytes (the bone marrow-derived cells or B cells) are those which secrete antibody. They also develop from haemopoietic stem cells, but their differentiation is not determined by the thymus. In birds, they are differentiated in an organ analogous to the thymus, called the Bursa of Fabricius. In mammals, however, no equivalent organ has been discovered, and it is thought that these B cells differentiate within the bone marrow.

It is now recognized that T cells are divided into at least several subtypes, termed "helper", "suppressor", and "killer" T cells, which have the function of (respectively) promoting a reaction, suppressing a reaction, or killing (lysing) foreign cells. These subclasses are well understood for murine systems, but they have only recently been described for human systems. See, for example, R. L. Evans, et al., *Journal of Experimental Medicine*, Volume 145, 221-232, 1977; and L. Chess and S. F. Schlossman-"Functional Analysis of Distinct Human T-Cell Subsets Bearing Unique Differentiation Antigens", in "Contemporary Topics in Immunobiology", O. Stutman, Editor, Plenum Press, 1977, Volume 7, 363-379.

The ability to identify or suppress classes or subclasses of T cells is important for diagnosis or treatment of various immunoregulatory disorders or conditions.

For example, certain leukemias and lymphomas have differing prognosis depending on whether they are of B cell or T cell origin. Thus, evaluation of the disease prognosis depends upon distinguishing between these two classes of lymphocytes. See, for example, A. C. Aisenberg and J. C. Long, *The American Journal of Medicine*, 58:300 (March, 1975); D. Belpomme, et al., in "Immunological Diagnosis of Leukemias and Lymphomas", S. Thierfelder, et al., eds, Springer, Heidelberg, 1977, 33-45; and D. Belpomme, et al., *British Journal of Haematology*, 1978, 38, 85.

Certain disease states (e.g., juvenile rheumatoid arthritis, certain leukemias, and agammaglobulinemia) are associated with an imbalance of T cell subclasses. It has been suggested that autoimmune diseases generally are associated with an excess of "helper" T cells or a deficiency of certain "suppressor" T cells, while agammaglobulinemia is associated with an excess of certain "suppressor" T cells or a deficiency of "helper" T cells. Malignancies generally are associated with an excess of "suppressor" T cells.

In certain leukemias, excess T cells are produced in an arrested stage of development. Diagnosis may thus depend on the ability to detect this imbalance or excess. See, for example, J. Kersey, et al., "Surface Markers Define Human Lymphoid Malignancies with Differing Prognoses" in *Haematology and Blood Transfusion*, Volume 20, Springer-Verlag, 1977, 17-24, and references contained therein.

Acquired agammaglobulinemia, a disease state in which no immune globulin is produced, comprises at least two distinct types. In type I the failure to produce immune globulin is due to an excess of suppressor T cells, while in type II it is due to a lack of helper T cells. In both types, there appears to be no defect or lack in

the patients' B cells, the lymphocytes which are responsible for the actual secretion of the antibody; however, these B cells are being either suppressed or "not helped", resulting in greatly decreased or absent immune globulin production. The type of acquired agammaglobulinemia may thus be determined by testing for an excess of suppressor T cells or an absence of helper T cells.

On the therapeutic side, there is some suggestion, as yet not definitely proven, that administration of antibodies against the subtype of T cell in excess may have therapeutic benefit in autoimmune disease or malignancies. For example, a helper T cell cancer (certain cutaneous T cell lymphomas and certain T cell acute lymphoblastic leukemias) may be treated by an antibody to a helper T cell antigen. Treatment of autoimmune disease caused by an excess of helper cells may also be accomplished in the same fashion.

Antisera against the entire class of human T cells (so-called antihuman thymocyte globulin or ATG) has been reported useful therapeutically in patients receiving organ transplants. Since the cell-mediated immune response (the mechanism whereby transplants are rejected) depends upon T cells, administration of antibody to T cells prevents or retards this rejection process. See, for example, Cosimi, et al., "Randomized Clinical Trial of ATG in Cadaver Renal Allgraft Recipients: Importance of T Cell Monitoring", *Surgery* 40:155-163 (1976) and references contained therein.

The identification and suppression of human T cell classes and subclasses has previously been accomplished by the use of spontaneous autoantibodies or selective antisera for human T cells obtained by immunizing animals with human T cells, bleeding the animals to obtain serum, and adsorbing the antiserum with (for example) autologous but not allogeneic B cells to remove antibodies with unwanted reactivities. The preparation of these antisera is extremely difficult, particularly in the adsorption and purification steps. Even the adsorbed and purified antisera contain many impurities in addition to the desired antibody, for several reasons. First, the serum contains millions of antibody molecules even before the T cell immunization. Second, the immunization causes production of antibodies against a variety of antigens found on all human T cells injected. There is no selective production of antibody against a single antigen. Third, the titer of specific antibody obtained by such methods is usually quite low, (e.g., inactive at dilutions greater than 1:100) and the ratio of specific to non-specific antibody is less than 1/10⁶.

See, for example, the Chess and Schlossman article referred to above (at pages 365 and following) and the Chemical and Engineering News article referred to above, where the deficiencies of prior art antisera and the advantages of monoclonal antibody are described.

SUMMARY OF THE INVENTION

There has now been discovered a novel hybridoma (designated OKT4) which is capable of producing novel monoclonal antibody against an antigen found on essentially all normal human peripheral helper T cells (about 55% of normal human peripheral T cells). The antibody so produced is monospecific for a single determinant on normal human helper T cells and contains essentially no other anti-human immunoglobulin, in contrast to prior art antisera (which are inherently contaminated with antibody reactive to numerous human antigens) and to prior art monoclonal antibodies (which

are not monospecific for a human helper T cell antigen). Moreover, this hybridoma can be cultured to produce antibody without the necessity of immunizing and killing animals, followed by the tedious adsorption and purification steps necessary to obtain even the impure antisera of the prior art. It is accordingly one object of this invention to provide hybridomas which produce antibodies against an antigen found on essentially all normal human helper T cells.

It is a further aspect of the present invention to provide methods for preparing these hybridomas.

A further object of the invention is to provide essentially homogeneous antibody against an antigen found on essentially all normal human helper T cells.

A still further object is to provide methods for treatment or diagnosis of disease employing these antibodies.

Other objects and advantages of the invention will become apparent from the examination of the present disclosure.

In satisfaction of the foregoing objects and advantages, there is provided by this invention a novel hybridoma producing novel antibody to an antigen found on essentially all normal human helper T cells, the antibody itself, and diagnostic and therapeutic methods employing the antibody. The hybridoma was prepared generally following the method of Milstein and Kohler. Following immunization of mice with normal E rosette positive human T cells, the spleen cells of the immunized mice were fused with cells from a mouse myeloma line and the resultant hybridomas screened for those with supernatants containing antibody which gave selective binding to normal E rosette positive human T cells. The desired hybridomas were subsequently cloned and characterized. As a result, a hybridoma was obtained which produces antibody (designated OKT4) against an antigen on essentially all normal human helper T cells. Not only does this antibody react with essentially all normal human peripheral helper T cells, but it also does not react with other normal peripheral blood lymphoid cells, including non-helper T cells. In addition, the cell surface antigen recognized by this antibody is detected on approximately 80% of normal human thymocytes. Type II acquired agammaglobulinemia patients were detected by OKT4 antibody in a blind test. The subject antibody also reacts with about 55% of Rhesus monkey peripheral T cells.

In view of the difficulties indicated in the prior art and the lack of success reported using malignant cell lines as the antigen, it was surprising that the present method provided the desired hybridoma. It should be emphasized that the unpredictable nature of hybrid cell preparation does not allow one to extrapolate from one antigen or cell system to another. In fact, the present applicants have discovered that using a T cell malignant cell line as the antigen caused formation of hybridomas which did not produce the desired antibody. Attempts to use purified antigens separated from the cell surfaces were also unsuccessful.

Both the subject hybridoma and the antibody produced thereby are identified herein by the designation "OKT4", the particular material referred to being apparent from the context. The subject hybridoma OKT4 was deposited in the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Md. 20852 on Apr. 26, 1979, and was given the ATCC accession number CRL 8002.

The preparation and characterization of the hybridoma and the resultant antibody will be better under-

stood by reference to the following description and Examples.

DETAILED DESCRIPTION OF THE INVENTION

The method of preparing the hybridoma generally comprises the following steps:

- A. Immunizing mice with E rosette positive purified normal human peripheral T cells. While it has been found that female CAF₁ mice are preferred, it is contemplated that other mouse strains could be used. The immunization schedule and T cell concentration should be such as to produce useful quantities of suitably primed splenocytes. Three immunizations at fourteen day intervals with 2×10^7 cells/mouse/injection in 0.2 ml phosphate buffered saline has been found to be effective.
- B. Removing the spleens from the immunized mice and making a spleen suspension in an appropriate medium. About one ml of medium per spleen is sufficient. These experimental techniques are well-known.
- C. Fusing the suspended spleen cells with mouse myeloma cells from a suitable cell line by the use of a suitable fusion promoter. The preferred ratio is about 5 spleen cells per myeloma cell. A total volume of about 0.5-1.0 ml of fusion medium is appropriate for about 10^8 splenocytes. Many mouse myeloma cell lines are known and available, generally from members of the academic community or various deposit banks, such as the Salk Institute Cell Distribution Center, La Jolla, CA. The cell line used should preferably be of the so-called "drug resistant" type, so that unfused myeloma cells will not survive in a selective medium, while hybrids will survive. The most common class is 8-azaguanine resistant cell lines, which lack the enzyme hypoxanthine guanine phosphoribosyl transferase and hence will not be supported by HAT (hypoxanthine, aminopterin, and thymidine) medium. It is also generally preferred that the myeloma cell line used be of the so-called "non-secreting" type, in that it does not itself produce any antibody, although secreting types may be used. In certain cases, however, secreting myeloma lines may be preferred. While the preferred fusion promoter is polyethylene glycol having an average molecular weight from about 1000 to about 4000 (commercially available as PEG 1000, etc.), other fusion promoters known in the art may be employed.
- D. Diluting and culturing in separate containers, the mixture of unfused spleen cells, unfused myeloma cells, and fused cells in a selective medium which will not support the unfused myeloma cells for a time sufficient to allow death of the unfused cells (about one week). The dilution may be a type of limiting one, in which the volume of diluent is statistically calculated to isolate a certain number of cells (e.g., 1-4) in each separate container (e.g., each well of a microtiter plate). The medium is one (e.g., HAT medium) which will not support the drug-resistant (e.g., 8-azaguanine resistant) unfused myeloma cell line. Hence, these myeloma cells perish. Since the unfused spleen cells are non-malignant, they have only a finite number of generations. Thus, after a certain period of time (about one week) these unfused spleen cells fail to reproduce. The fused cells, on the other hand, continue

to reproduce because they possess the malignant quality of the myeloma parent and the ability to survive in the selective medium of the spleen cell parent.

- E. Evaluating the supernatant in each container (well) containing a hybridoma for the presence of antibody to E rosette positive purified human T cells.
- F. Selecting (e.g., by limiting dilution) and cloning hybridomas producing the desired antibody.

Once the desired hybridoma has been selected and cloned, the resultant antibody may be produced in one of two ways. The purest monoclonal antibody is produced by in vitro culturing of the desired hybridoma in a suitable medium for a suitable length of time, followed by recovery of the desired antibody from the supernatant. The suitable medium and suitable length of culturing time are known or are readily determined. This in vitro technique produces essentially monospecific monoclonal antibody, essentially free from other specific antihuman immune globulin. There is a small amount of other immune globulin present since the medium contains xenogeneic serum (e.g. fetal calf serum). However, this in vitro method may not produce a sufficient quantity or concentration of antibody for some purposes, since the concentration of monoclonal antibody is only about 50 µg/ml.

To produce a much greater concentration of slightly less pure monoclonal antibody, the desired hybridoma may be injected into mice, preferably syngenic or semi-syngenic mice. The hybridoma will cause formation of antibody-producing tumors after a suitable incubation time, which will result in a high concentration of the desired antibody (about 5-20 mg/ml) in the bloodstream and peritoneal exudate (ascites) of the host mouse. Although these host mice also have normal antibodies in their blood and ascites, the concentration of these normal antibodies is only about 5% of the monoclonal antibody concentration. Moreover, since these normal antibodies are not antihuman in their specificity, the monoclonal antibody obtained from the harvested ascites or from the serum is essentially free of any contaminating antihuman immune globulin. This monoclonal antibody is high titer (active at dilutions of 1:50,000 or higher) and high ratio of specific to non-specific immune globulin (about 1/20). Immune globulin produced incorporating the κ light myeloma chains are non-specific, "nonsense" peptides which merely dilute the monoclonal antibody without detracting from its specificity.

EXAMPLE I

Production of Monoclonal Antibodies

A. Immunization and Somatic Cell Hybridization

Female CAF₁ mice (Jackson Laboratories; 6-8 weeks old) were immunized intraperitoneally with 2×10^7 E rosette purified T cells in 0.2 ml of phosphate buffered saline at 14-day intervals. Four days after the third immunization, spleens were removed from the mice, and a single cell suspension was made by pressing the tissue through a stainless steel mesh.

Cell fusion was carried out according to the procedure developed by Kohler and Milstein. 1×10^8 splenocytes were fused in 0.5 ml of a fusion medium comprising 35% polyethylene glycol (PEG 1000) and 5% dimethylsulfoxide in RPMI 1640 medium (Gibco, Grand Island, NY) with 2×10^7 P3X63Ag8U1 myeloma cells

supplied by Dr. M. Scharff, Albert Einstein College of Medicine, Bronx, NY. These myeloma cells secrete IgG₁ κ light chains.

B. Selection and Growth of Hybridoma

After cell fusion, cells were cultured in HAT medium (hypoxanthine, aminopterin, and thymidine) at 37° C. with 5% CO₂ in a humid atmosphere. Several weeks later, 40 to 100 μ l of supernatant from cultures containing hybridomas were added to a pellet of 10⁶ peripheral lymphocytes separated into E rosette positive (E⁺) and E rosette negative (E⁻) populations, which were prepared from blood of healthy human donors as described by Mendes (*J. Immunol.* 111:860, 1973). Detection of mouse hybridoma antibodies binding to these cells was determined by radioimmunoassay and/or indirect immunofluorescence. In the first method, the cells were initially reacted with 100 μ l of affinity-purified ¹²⁵I goat-anti-mouse IgG (10⁶ cpm/ μ g; 500 μ g/ μ l). (Details of iodination of goat-anti-mouse IgG were described by Kung, et al., *J. Biol. Chem.* 251(8): 2399, 1976). Alternatively, cells incubated with culture supernatants were stained with a fluoresceinated goat-anti-mouse IgG (G/M FITC) (Meloy Laboratories, Springfield, VA; F/p=2.5) and the fluorescent antibody-coated cells were subsequently analyzed on the Cytofluorograf FC200/4800A (Ortho Instruments, Westwood, MA) as described in Example III. Hybridoma cultures containing antibodies reacting specifically with E⁺ lymphocytes (T cells) were selected and cloned. Subsequently, the clones were transferred intraperitoneally by injecting 1 \times 10⁷ cells of a given clone (0.2 ml volume) into CAF₁ mice primed with 2,6,10,14-tetramethylpentadecane, sold by Aldrich Chemical Company under the name Pristine. The malignant ascites from these mice were then used to characterize lymphocytes as described below in Example II. The subject hybrid antibody OKT4 was demonstrated by standard techniques to be of IgG₂ subclass and to fix complement.

EXAMPLE II

Characterization of OKT4 Reactivity

A. Isolation of Lymphocyte Populations

Human peripheral blood mononuclear cells were isolated from healthy volunteer donors (ages 15-40) by Ficoll-Hypaque density gradient centrifugation (Pharmacia Fine Chemicals, Piscataway, NJ) following the technique of Boyum, *Scand. J. Clin. Lab. Invest.* 21 (Suppl. 97): 77, 1968. Unfractionated mononuclear cells were separated into surface Ig⁺ (B) and Ig⁻ (T plus Null) populations by Sephadex G-200 anti-F(ab')₂ column chromatography as previously described by Chess, et al., *J. Immunol.* 113: 1113 (1974). T cells were recovered by E rosetting the Ig⁻ population with 5% sheep erythrocytes (Microbiological Associates, Bethesda, MD). The rosetted mixture was layered over Ficoll-Hypaque and the recovered E⁺ pellet treated with 0.155 M NH₄Cl (10 ml per 10⁸ cells). The T cell population so obtained was <2% EAC rosette positive and >95% E rosette positive as determined by standard methods. In addition, the non-rosetting Ig⁻ (Null cell) population was harvested from the Ficoll interface. This latter population was <5% E⁺ and \leq 2% sIg⁺. The surface Ig⁺ (B) population was obtained from the Sephadex G-200 column following elution with normal human gamma globulin as previously described. This population was >95% surface Ig⁺ and <5% E⁺.

Normal human macrophages were obtained from the mononuclear population by adherence to polystyrene. Thus, mononuclear cells were resuspended in final culture media (RPMI 1640, 2.5 mM HEPES [4-(2-hydroxyethyl)-1-piperazinepropane sulfonic acid] buffer, 0.5% sodium bicarbonate, 200 mM L-glutamine, and 1% penicillin-streptomycin, supplemented with 20% heat-inactivated human AB serum) at a concentration of 2 \times 10⁶ cells and incubated in plastic petri dishes (100 \times 20 mm) (Falcon Tissue Culture Dish; Falcon, Oxnard, CA) at 37° C. overnight. After extensive washing to remove non-adherent cells, the adherent population was detached by brisk washing with cold serum-free medium containing 2.5 mM EDTA and occasional scraping with the rubber tip of a disposable syringe plunger. Greater than 85% of the cell population was capable of ingesting latex particles and had morphologic characteristics of monocytes by Wright-Giemsa staining.

B. Normal Thymus

Normal human thymus gland was obtained from patients aged two months to 14 years undergoing corrective cardiac surgery. Freshly obtained portions of the thymus gland were immediately placed in 5% fetal calf serum in medium 199 (Gibco), finely minced with forceps and scissors, and subsequently made into single cell suspensions by being pressed through wire mesh. The cells were next layered over Ficoll-Hypaque and spun and washed as previously described in section A above. The thymocytes so obtained were >95% viable and \geq 90% E rosette positive.

C. Cell Lines

Epstein-Barr Virus (EBV) transformed B cell lines from four normal individuals (Laz 007, Laz 156, Laz 256, and SB) and T cell lines CEM, HJD-1, Laz 191, and HM1 established from leukemic patients were provided by Dr. H. Lazarus, Sidney Farber Cancer Institute, Boston, MA.

D. T Acute Lymphoblastic Leukemia (T-ALL) Cells and T Chronic Lymphatic Leukemia (T-CLL) Cells

Leukemia cells were obtained from 12 patients with T-ALL. These individuals' cells had previously been determined to be of T cell lineage by their spontaneous rosette formation with sheep erythrocytes (>20% E⁺) and reactivity with T cell specific hetero-antisera, anti-HTL (anti-B.K.) and A99, as previously described by Schlossman, et al., *Proc. Nat. Acad. Sci.* 73: 1288 (1976). Tumor cells from three individuals were reactive (TH₂⁺) with rabbit and/or equine anti-TH₂ while cells from the remaining nine were non-reactive (TH₂⁻). Leukemic cells from two patients with TH₂⁻ T-CLL were also utilized. Both acute and chronic T cell leukemia cells were cryopreserved in -196° C. vapor phase liquid nitrogen in 10% dimethylsulfoxide and 20% AB human serum until the time of surface characterization. The tumor populations analyzed were >90% blasts by Wright-Giemsa morphology in all instances.

EXAMPLE III

Cytofluorographic Analysis and Cell Separation

Cytofluorographic analysis of all cell populations was performed by indirect immunofluorescence with fluorescein-conjugated goat-anti-mouse IgG (G/M FITC) (Meloy Laboratories) on a Cytofluorograf

FC200/4800A (Ortho Instruments). In brief, $1-2 \times 10^6$ cells were treated with 0.15 ml OKT4 at a 1:1000 dilution, incubated at 4° C. for 30 minutes, and washed twice. The cells were then reacted with 0.15 ml of a 1:40 dilution G/M FITC at 4° C. for 30 minutes, centrifuged, and washed three times. These cells were then analyzed on the Cytofluorograf and the intensity of fluorescence per cell recorded on a pulse height analyzer. A similar pattern of reactivity was observed at a dilution of 1:50,000, but further dilution caused loss of reactivity. Background staining was obtained by substituting a 0.15 ml aliquot of 1:1000 ascites from a Balb/cJ mouse intraperitoneally immunized with a non-producing hybrid clone.

In experiments designed to separate OKT4+ and OKT4- cells, 100×10^6 unfractionated mononuclear cells or thymocytes were labeled with 4 ml of a 1:1000 dilution of OKT4 and developed with G/M FITC. An identical staining approach was utilized to prepare human T cells isolated as in Example IIA above. Utilizing a fluorescence activated cell sorter (FACS-I) (Becton-Dickinson, Mountain View, CA), lymphocytes were separated into OKT4+ and OKT4- populations. Post sort viability was >95% by Trypan blue exclusion in all instances. Purity of all separated populations was $\geq 95\%$.

EXAMPLE IV

Analysis of FACS Separated OKT4+ and OKT4- Subsets with Equine Anti-TH₂

OKT4+ and OKT4- T cells were separated on FACS and placed in culture at 2×10^6 cells per ml in RPMI 1640 (Grand Island Biological Company), containing 20% human AB serum, 1% penicillin-streptomycin, 200 mM L-glutamine, 25 mM HEPES buffer (Microbiological Associates), and 0.5% sodium bicarbonate. After 24 hours in a 5% CO₂ humid atmosphere at 37° C., $1-2 \times 10^6$ cells of each population were reacted with equine anti-TH₂ and stained with fluorescein-conjugated IgG fraction rabbit anti-horse Ig (Cappel Laboratories, Downingtown, PA) as described by Reinherz and Schlossman, *J. Immunol.* 122: 1335-1341 (1979). Background staining was determined by substituting normal horse IgG for specific antibody and staining as above.

EXAMPLE V

Functional Studies

A. Proliferative Studies

The mitogenic response of the unseparated and FACS-fractionated lymphoid cells was tested in microculture to optimal and suboptimal doses of Concanavalin A (Con A) (Calbiochem, La Jolla, CA) and Phytohemagglutinin (PHA) (Burroughs-Wellcome Company, Greenville, NC). Alloantigen proliferative response was measured concurrently for these same populations using mitomycin c treated Laz 156, an EBV transformed human B lymphoblastoid cell line, as a stimulus. Proliferation to tetanus toxoid (Massachusetts Department of Public Health Biological Laboratories, Boston, MA) was tested utilizing 10 µg/ml final concentration. Herpes-Zoster antigen was kindly provided by Dr. John Zaia (Harvard Medical School, Boston, MA) and utilized at a 1:6 dilution. Five percent macrophages obtained in the manner described above were added to all populations at the initiation of in vitro cultures. Mitogen stimulated cultures were pulsed after four days with

0.2 µCi of ³H-thymidine (³H-TdR) (1.9 Ci/mM specific activity) (Schwartz-Mann, Division of Becton, Dickinson, Orangeburg, NY) and harvested 18 hours later on a MASH II apparatus (Microbiological Associates). ³H-TdR incorporation was measured in a Packard Scintillation Counter (Packard Instrument Company, Downer's Grove, IL). Background ³H-TdR incorporation was obtained by substituting media for mitogen. Soluble antigen and alloantigen stimulated cultures were pulsed after five days with ³H-TdR for 18 hours, harvested, and counted as above.

B. Cytotoxicity Studies

Sensitization cultures for cell-mediated lympholysis (CML) were established by placing unfractionated T cells, FACS-separated OKT4+ and OKT4- T cell subsets, or different ratios of recombined OKT4+ and OKT4- T cells with mitomycin-treated stimulator cells all at 2×10^6 cells per ml in multiple microtitre plate wells. At the end of five days, non-viable cells were removed by Ficoll-Hypaque centrifugation. These unfractionated and fractionated T cell populations were then added to ⁵¹Cr sodium chromate labeled target cells and specific chromium release determined after a six-hour cell incubation. In other experiments, unfractionated T cells were sensitized with mitomycin-treated stimulator cells as above and then fractionated into OKT4+ and OKT4- T cell subsets on FACS after five days in MLC and specific chromium release determined. Percent cytotoxicity was determined with the following formula:

$$\frac{^{51}\text{Cr released by experiment} - ^{51}\text{Cr released spontaneously}}{^{51}\text{Cr released by freeze-thaw} - ^{51}\text{Cr released spontaneously}} \times 100$$

All samples were performed in triplicate and results expressed as the mean. Spontaneous release was less than 20% of maximal lysis in all cases.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows the fluorescence pattern obtained on the Cytofluorograf after reacting normal human peripheral T cells with OKT4 at a 1:1000 dilution and G/M FITC. For comparison, results with monoclonal antibodies OKT1 and OKT3 are shown under equivalent conditions in FIGS. 1-5.

FIG. 2 shows the fluorescence pattern obtained on the Cytofluorograf after reacting human thymocytes with OKT4 and G/M FITC.

FIG. 3 shows the fluorescence pattern obtained on the Cytofluorograf after reacting leukemic cells from B cell chronic lymphoblastic leukemia patients with OKT4 and G/M FITC.

FIG. 4 shows the fluorescence pattern obtained on the Cytofluorograf after reacting the human T cell line HJD-1 with OKT4 and G/M FITC.

FIG. 5 shows the fluorescence pattern obtained on the Cytofluorograf after reacting the human T cell line CEM with OKT4 and G/M FITC.

FIG. 6 shows the results of reaction of T cell populations with equine anti-TH₂ serum.

FIG. 7 shows the cytotoxic capacity of unfractionated T cells and T cell subsets. Percent specific lysis is shown on the ordinate and effector/target (E/T) ratio is shown on the abscissa.

The production of the hybridoma and the production and characterization of the resulting monoclonal antibody were conducted as described in the above Examples. Although large quantities of the subject antibody were prepared by injecting the subject hybridoma intraperitoneally into mice and harvesting the malignant ascites, it is clearly contemplated that the hybridoma could be cultured *in vitro* by techniques well-known in the art and the antibody removed from the supernatant. As shown in FIG. 1, approximately 45% of the human peripheral blood T cell population of a given normal individual is reactive with OKT4, whereas the entire B cell, null cell, and macrophage populations isolated from the same individual are unreactive with OKT4. Similar results were obtained on populations of lymphocytes from fifteen other normal individuals. The monoclonal antibody is thus characterized in that it is reactive with an antigen contained on the surface of approximately 55% of normal human peripheral T cells, while being unreactive with any antigens on the surface of the other three cell types discussed above. As will be discussed below, the OKT4⁺ portion of the human peripheral T cell population is the subclass of helper T cells. This differential reactivity is one test by which the subject antibody OKT4 may be detected and distinguished from other antibodies.

As shown in FIG. 2, approximately 80% of normal human thymocytes from a six-month old infant are reactive with OKT4. Similar results (about 80% reactivity) were obtained using six additional thymus specimens from normal individuals two months to 19 years of age. The pattern of reactivity in FIG. 2 provides a second method of detecting the subject antibody OKT4 and distinguishing it from other antibodies.

As shown in FIG. 3, the subject antibody is unreactive with leukemic cells from B cell chronic lymphoblastic leukemia.

A further characterization of the subject antibody OKT4 is shown by the reactivity to various human T cell lines illustrated in FIGS. 4 and 5. As can be seen, the reactivity of the subject antigen to human T cell lines was heterogeneous, being strong for the line CEM, and nonexistent for the line HDJ-1. There was also no reactivity with the cell lines Laz 191 and HM1. This differential reactivity of OKT4 to various readily-available human T cell lines provides yet another method of characterizing and describing the subject antibody.

FIG. 6 illustrates the reactivity of OKT4 separated subsets with anti-TH₂. About 25% of the unfractionated T cell population is reactive with anti-TH₂. In contrast, the OKT4⁺ population contains no cells which are reactive with anti-TH₂, while the OKT4⁻ population is predominantly TH₂⁺ and contains all the TH₂⁺ cells found in the unfractionated T cell population. This indicated that the TH₂⁺ and the OKT4⁺ subsets are reciprocal and distinct from one another.

FIG. 7 illustrates the cytotoxic capacity of unfractionated, OKT4⁺, and OKT4⁻ T cells. The OKT4⁺ population is only minimally cytotoxic, while the degree of killing mediated by the OKT4⁻ population is greater than for the unfractionated T cell population. The differential reactivities of OKT4⁺ and OKT4⁻ T cell populations shown in FIGS. 6 and 7 provide a further means of characterizing the subject antibody.

The results illustrated in FIGS. 1-5 are summarized and augmented with additional data in Table I below. The Table compares monoclonal antibodies produced by hybridomas designated OKT1, OKT3, and OKT4

(which last is the subject of the present application). In addition to the data in FIGS. 1-5, Table I also demonstrates that OKT4 is unreactive with normal human peripheral B cells, null cells and macrophages, as well as leukemic cells from T cell and null cell acute lymphoblastic leukemia patients and EBV transformed B cell lines. In contrast to OKT1, OKT4 does not react with leukemic cells from T cell chronic lymphoblastic leukemia patients.

Functional studies were conducted on lymphoid populations which had been separated on a fluorescence activated cell separator (FACS). The results of these studies are shown in Tables II through IV below and provide further support for the previously-described characterization of the subject monoclonal antibody.

In these studies, an unfractionated T cell population was treated with a 1:1000 dilution of OKT4 and G/M FITC and separated on the FACS in OKT4⁺ and OKT4⁻ subsets. Given the purity of the populations obtained (greater than or equal to 95%), 5% macrophages were added to the separated populations prior to *in vitro* culture. The unfractionated T cell population and isolated OKT4⁺ and OKT4⁻ T cell subsets were then stimulated with PHA, Con A, soluble antigens, and alloantigens to assess their *in vitro* proliferative responses.

The proliferative response of the unfractionated T cell populations to PHA and Con A is shown in Table II. A maximal proliferative response by the unfractionated T cell population is obtained with 1 µg of PHA per 10⁶ cells with diminished responses occurring at 0.5 µg and 0.1 µg of PHA per 10⁶ cells. Treatment of the unfractionated T cells with OKT4 and goat-mouse FITC without subsequent fractionation did not alter the proliferative response. In contrast, differences in response to PHA were obtained with the separated OKT4⁺ and OKT4⁻ T cell subset. The OKT4⁺ population of cells responded to all doses of PHA in a fashion similar to the unseparated T cell population. However, the proliferative response of OKT4⁻ cells was significantly less at all doses of PHA tested. Furthermore, at a dose of PHA of 0.1 µg per 10⁶ cells, the OKT4⁻ T cells did not proliferate at all, whereas the OKT4⁺ T cell subset and unfractionated cells were still responsive. The proliferative response of these subsets to Con A, on the other hand, was similar and the two subsets of cells could not be distinguished from one another or the unfractionated T cell population. The responses to alloantigen in MLC and to soluble antigens were next examined. As shown in Table III, the unfractionated T cell population, the unfractionated T cell population treated with OKT4 and G/M FITC, and both the OKT4⁺ and OKT4⁻ T cell subsets responded in a similar fashion in MLC against Laz 156. It should be noted, however, that the OKT4⁻ cells from two of six individuals tested, although proliferating significantly in MLC, incorporated less ³H-TdR than their respective OKT4⁺ subset (data not shown). In contrast, proliferative responses to soluble antigens provided the clearest distinction between the subsets. In all cases tested, the OKT4⁺ T cell subset proliferated well to the soluble antigens, tetanus toxoid and Herpes-Zoster, whereas the OKT4⁻ T cell subset was virtually unresponsive.

Table IV shows that the OKT4⁻ subset of T cells cannot generate much, if any, cytotoxicity when it is sensitized alone in MLC. Thus, although the OKT4⁻ T cells became cytotoxic/effectors in the unfractionated allo-sensitized T cell population, by itself it could not be

TABLE II-continued

| Proliferative Stimulus | MITOGEN RESPONSIVENESS OF UNFRACTIONATED AND MONOCLONAL ANTIBODY SEPARATED T CELL SUBSETS | | | |
|--------------------------------------|---|--|-----------------|---------------------------|
| | T Cells | T Cells Treated with OKT4 and G/M FITC | | OKT4 ⁻ T Cells |
| PHA (0.1 µg/10 ⁶ cells) | 4,215 ± 386 | 4,317 ± 523 | 4,999 ± 677 | 72 ± 11 |
| Con A (250 µg/10 ⁶ cells) | 199,310 ± 14,317 | 192,158 ± 14,258 | 159,560 ± 2,619 | 174,992 ± 20,179 |
| Con A (125 µg/10 ⁶ cells) | 100,726 ± 8,864 | 92,633 ± 6,780 | 97,003 ± 9,089 | 82,199 ± 6,154 |
| Con A (50 µg/10 ⁶ cells) | 50,673 ± 6,155 | 52,317 ± 7,813 | 49,329 ± 4,110 | 38,842 ± 6,735 |
| Media Control | 63 ± 2 | 57 ± 6 | 167 ± 8 | 69 ± 12 |

TABLE III

| PROLIFERATIVE RESPONSE OF UNFRACTIONATED T CELLS AND MONOCLONAL ANTIBODY SEPARATED T CELL SUBSETS TO SOLUBLE ANTIGENS AND ALLOANTIGENS | | | | |
|--|-----------------|--|-----------------|-----------------|
| Proliferative Stimulus | T Cells | T Cells Treated with OKT4 and G/M FITC | | |
| | | OKT4+ T Cells | OKT4- T Cells | |
| Experiment #1 | | | | |
| Laz 156 _m | 150,304 ± 6,966 | 149,810 ± 7,630 | 173,061 ± 4,336 | 167,087 ± 5,228 |
| Tetanus Toxoid | 18,046 ± 271 | 19,947 ± 3,214 | 19,660 ± 2,348 | 516 ± 69 |
| Herpes-Zoster | 47,413 ± 3,208 | 40,774 ± 4,305 | 55,785 ± 10,608 | 188 ± 129 |
| Media | 166 ± 22 | 180 ± 35 | 220 ± 10 | 106 ± 13 |
| Experiment #2 | | | | |
| Laz 156 _m | 67,265 ± 3,466 | 68,399 ± 6,154 | 97,615 ± 4,361 | 67,483 ± 4,792 |
| Tetanus Toxoid | 16,853 ± 1,033 | 18,597 ± 1,643 | 17,222 ± 911 | 206 ± 23 |
| Herpes-Zoster | 23,629 ± 3,677 | 22,889 ± 1,956 | 25,951 ± 2,609 | 169 ± 25 |
| Media | 167 ± 43 | 289 ± 57 | 298 ± 41 | 31 ± 8 |

TABLE IV

| Responder Population | EVIDENCE FOR T-T INTERACTIONS IN THE GENERATION OF MAXIMAL CYCOTOTOXICITY IN CML | | | |
|---|--|-------------------------------|---------------|------|
| | E/T Ratio: 20:1 | % Specific Lysis ⁺ | | |
| | | Experiment #1 | Experiment #2 | |
| Sensitized to Laz 156 _m | | | | |
| Unfractionated T Cells | 40+ | — | 23 | 11.9 |
| Unfractionated T Cells Treated with OKT4 + G/M FITC | | 44 | 24 | 25 |
| OKT4 ⁻ T Cells | | 6 | 3 | 10.9 |
| OKT4 ⁺ T Cells | | 26 | 14 | 18.3 |
| OKT4 ⁺ and OKT4 ⁻ T Cells ⁺⁺ | | 48 | 31 | 31 |

⁺S.R. less than 20% in all cases and S.D. less than or equal to 10% at all E/T ratios. Lysis by unsensitized T cells was less than or equal to 3% for all populations tested.

⁺⁺OKT4⁺ and OKT4⁻ recombined at 1:1 ratios.

What is claimed is:

1. A monoclonal antibody of class IgG produced by a hybridoma formed by fusion of cells from a mouse myeloma line and spleen cells from a mouse previously immunized with human T cells, which antibody:

- reacts with essentially all normal human peripheral helper T cells (being about 55% of all normal human peripheral T cells), but not with normal human peripheral B cells, null cells or macrophages;
- reacts with about 80% of normal human thymocytes;
- does not react with leukemic cells from humans with T cell chronic lymphoblastic leukemia, B cell chronic lymphoblastic leukemia, T cell acute lymphoblastic leukemia, or null cell acute lymphoblastic leukemia;

(d) reacts with the human T cell line CEM, but not with HJD-1, Laz 191, or HMI1;

(e) does not react with Epstein-Barr virus-transformed human B cell lines Laz 007, Laz 156, Laz 256, or SB;

(f) reacts with about 55% of Rhesus monkey peripheral T cells;

(g) fixes complement; and

(h) defines a T cell population which is unreactive with anti-TH₂ serum and is only minimally cytotoxic.

2. The monoclonal antibody of claim 1 which is of subclass IgG₂.

3. The monoclonal antibody of claim 1 which is produced from a hybridoma formed by fusion of P3X63Ag-8U1 myeloma cells and spleen cells from a CAF₁ mouse previously immunized with E rosette purified human T cells.

4. Mouse monoclonal antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages.

5. A method of preparing monoclonal antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages, which comprises culturing the hybridoma ATCC CRL 8002 in a suitable medium and recovering the antibody from the supernatant above said hybridoma.

6. The monoclonal antibody prepared by the method of claim 5.

7. A method of preparing monoclonal antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages, which comprises injecting into a mouse the hybridoma ATCC

17

CRL 8002 and recovering the antibody from the malignant ascites or serum of said mouse.

8. The monoclonal antibody prepared by the method of claim 7.

9. A monoclonal antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages prepared by the method, which comprises the steps of:

- (i) immunizing mice with E rosette positive purified human T cells;
- (ii) removing the spleens from said mice and making a suspension of the spleen cells;
- (iii) fusing said spleen cells with mouse myeloma cells in the presence of a fusion promoter;
- (iv) diluting and culturing the fused cells in separate wells in a medium which will not support the unfused myeloma cells;
- (v) evaluating the supernatant in each well containing a hybridoma for the presence of antibody to E rosette positive purified T cells;
- (vi) selecting and cloning a hybridoma producing antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages; and
- (vii) recovering the antibody from the supernatant above said clones.

10. A monoclonal antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or

18

macrophages prepared by the method, which comprises the steps of:

- (i) immunizing mice with E rosette positive purified human T cells;
- (ii) removing the spleens from said mice and making a suspension of the spleen cells;
- (iii) fusing said spleen cells with mouse myeloma cells in the presence of a fusion promoter;
- (iv) diluting and culturing the fused cells in separate wells in a medium which will not support the unfused myeloma cells;
- (v) evaluating the supernatant in each well containing a hybridoma for the presence of antibody to E rosette positive purified T cells;
- (vi) selecting and cloning a hybridoma producing antibody which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages;
- (vii) transferring said clones intraperitoneally into mice; and
- (viii) harvesting the malignant ascites or serum from said mice, which ascites or serum contains the desired antibody.

11. A monoclonal antibody of class IgG produced by a hybridoma formed by fusion of cells from a mouse myeloma line and spleen cells from a mouse previously immunized with human T cells which reacts with essentially all normal human peripheral helper T cells but not with normal human peripheral B cells, null cells, or macrophages.

* * * * *

35

40

45

50

55

60

65